

1 **Title:** Effective microorganism – x attenuates circulating superoxide dismutase following an
2 acute bout of intermittent running in hot, humid conditions.

3
4 **Running title:** Effective microorganism x attenuates repeated sprint induced disturbances to
5 redox balance.

6
7 **Key Words:** Heat shock proteins (HSP), oxidative stress, redox balance, repeated sprint,
8 exercise, HIT, human.

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38 **Abstract**

39 This study determined the effectiveness of antioxidant supplementation on high intensity
40 exercise-heat stress.

41 Six males completed a high intensity running protocol twice in temperate conditions (TEMP;
42 20.4°C), and twice in hot conditions (HOT; 34.7°C). Trials were completed following seven
43 days supplementation with 70mL.day⁻¹ effective microorganism-x (EM-X; TEMP_{EMX} or
44 HOT_{EMX}) or placebo (TEMP_{PLA} or HOT_{PLA}). Plasma extracellular Hsp72 (eHsp72) and
45 superoxide dismutase (SOD) were measured by ELISA.

46 eHsp72 and SOD increased pre-post exercise ($p < 0.001$), with greater eHsp72 ($p < 0.001$)
47 increases observed in HOT (+1.5ng.mL⁻¹) compared to TEMP (+0.8ng.mL⁻¹). EM-X did not
48 influence eHsp72 ($p > 0.05$). Greater ($p < 0.001$) SOD increases were observed in HOT
49 (+0.22U.mL⁻¹) vs. TEMP (+0.10U.mL⁻¹) with SOD reduced in HOT_{EMX} vs. HOT_{PLA}
50 ($p = 0.001$). Physiological and perceptual responses were all greater ($p < 0.001$) in HOT vs.
51 TEMP conditions, with no difference followed EM-X ($p > 0.05$).

52 EM-X supplementation attenuated the SOD increases following HOT, potentiating its
53 application as an ergogenic aid to ameliorate oxidative stress.

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60 **Introduction**

61 Increased extracellular heat shock protein 72 (eHsp72) concentrations are evident in response
62 to exercise (Yamada, Amorim, Moseley, & Schneider, 2008) with the greater increases
63 during exercise-heat stress predicted by the magnitude of change in rectal temperature
64 (Gibson et al., 2014) and exercise intensity (Periard, Ruell, Caillaud, & Thompson, 2012).

65 Reactive oxygen species generation and/or disturbances to redox balance are essential
66 components of adaption (Powers, Duarte, Kavazis, & Talbert, 2010). Conversely, their
67 presence can induce performance decrements when repeated demanding exercise is required
68 with insufficient recovery (Cobley, McGlory, Morton, & Close, 2011) i.e. during tournament
69 competition or within consecutive day, multiday events. Oxidative stress can inhibit the onset
70 of tissue repair (Ascensao et al., 2008) with chronic oxidative stress coinciding with skeletal
71 muscle atrophy (Powers, et al., 2010). Amelioration of short-term negative effects of
72 oxidative stress on demanding exercise with limited recovery time could be advantageous,
73 thus this paradigm (exercise-heat stress; redox balance; Hsp72) requires exploration. N-
74 Acetylcysteine supplementation can achieve this goal, though nausea and gastrointestinal
75 discomfort can present (Cobley, et al., 2011), reducing its practicality. At present, there is no
76 literature directly assessing the interplay between oxidative stress/redox balance disturbances
77 and eHsp72 expression in vivo, though redox disturbances preceding increases in intracellular
78 Hsp72 (iHsp72) are well reported (Taylor et al., 2012).

79 Antioxidant supplementation may hypothetically blunt the exercise induced eHsp72 response
80 in a similar manner to that seen with hypoxic preconditioning and iHsp72 (Taylor, et al.,
81 2012), in addition to antioxidant mediated ergogenicity specific to high-intensity interval
82 training (HIT) type exercise (Cobley, et al., 2011). Effective Microorganism X (EM-X) has
83 potent antioxidant effects (Deiana et al., 2002; Do, Seo, Hwang, Kim, & Nam, 2007). The
84 antioxidant cocktail (EM-X) is derived from effective microorganisms of lactic acid bacteria,
85 yeast and photosynthetic bacteria (Aruoma et al., 2002) and presents no mutagenic effects

86 under chronic or acute supplementation (Ke, Liang, Zhong, Higa, & Aruoma, 2005). EM-X
87 administration increases serum dismutase, decreases malondialdehyde (Deiana, et al., 2002),
88 whilst, resisting acute severe oxidative stress mediated damage in the kidney and liver of rats
89 (Aruoma, et al., 2002), and conveying an anti-inflammatory influence at a cellular level,
90 independent of antioxidant activity (Do, et al., 2007). Given the potent stimuli that exercise
91 (Yamada, et al., 2008), exercise-heat stress (Gibson, et al., 2014) and disturbances to redox
92 (Taylor, et al., 2012) represent for HSP induction, supplementation of EM-X may affect
93 exercise induced disturbances to redox balance, thus the HSP exercise response. HIT specific
94 exercise-heat stress induced fatigue, as shown elsewhere with N-Acetylcysteine (Cobley, et
95 al., 2011), may be attenuated with EM-X supplementation by reducing disturbances to redox
96 balance (Aruoma, et al., 2002; Aruoma et al., 2003; Deiana, et al., 2002) and pro-
97 inflammatory cascades (Do, et al., 2007).

98 The aims of the present study are to investigate; i) the influence of HIT on eHsp72
99 concentration and plasma superoxide dismutase (SOD) activity; ii) the influence of ambient
100 temperature (thermoneutral/hyperthermic) on any HIT induced alterations in basal eHsp72
101 concentration and plasma SOD activity; iii) the influence of EM-X supplementation on HIT
102 induced eHsp72 concentration and plasma SOD activity within both environmental
103 conditions (thermoneutral/hyperthermic).

104 **Methods**

105 Subjects and general experimental controls/methods:

106 Six male subjects (mean \pm SD: age 22.0 ± 1.3 years; height 181.0 ± 4.19 cm; mass 73.5 ± 3.1
107 kg; maximum oxygen uptake (VO_{2max}) 51.7 ± 7 mL \cdot kg $^{-1}$ \cdot min $^{-1}$) volunteered to participate
108 within the present study. Subjects attended the laboratory at the same time of day to minimise
109 circadian variation on performance (Reilly et al., 2007; Winget, Deroshia, & Holley, 1985).

110 The confounding variables of hypoxic, thermal and hyperbaric exposures, and smoking,
111 glutamine, caffeine, alcohol and generic supplementation were all controlled in line with
112 previous HSP/redox balance exercise projects within the field (Taylor, et al., 2012).

113 Prior to reporting to the laboratory, subjects were instructed to drink 500 mL of water 2 hrs
114 prior to all laboratory visits, in accordance with the ACSM position stand (Sawka et al.,
115 2007). Upon arrival at the laboratory the subjects provided a urine sample for assessment of
116 specific gravity to determine hydration status using dip test strips (Combur10-test, Roche
117 Diagnostics, Mannheim, Germany). Nude body mass was assessed (Tanita BWB-800, Tokyo,
118 Japan), a rectal temperature probe was inserted at a depth of 10 cm past the anal sphincter
119 (Libra Medical, Reading, UK) and a HR monitor was affixed to the chest (Polar Sports
120 Tester, Polar Electro Oy, Kempele, Finland).

121 *High intensity interval running protocol (HIT_{RP}):* The protocol consisted of 20 fast high
122 intensity runs 10 seconds in duration at a velocity corresponding to the final running speed
123 achieved during the Maximal Anaerobic Running Test (MART) test ($23.0 \pm 1.8 \text{ km.h}^{-1}$). with
124 80 seconds of active recovery at a velocity corresponding to 35% $\text{VO}_{2\text{max}}$ ($6.7 \pm 2.2 \text{ km.h}^{-1}$)
125 and lasted approximately 40 minutes (37.3 ± 3.5 minutes). The high intensity running speed
126 was supramaximal in relation to the $\text{VO}_{2\text{max}}$ test. Heart rate (HR) was recorded following
127 each 10 second run and 60 seconds into recovery. Ratings of perceived exertion (RPE; (Borg,
128 Ljunggren, & Ceci, 1985)), thermal sensation (TS; (Gagge, Stolwijk, & Saltin, 1969)) and
129 core temperature (T_c) were recorded 60 seconds into recovery. Venous whole blood samples
130 were obtained at rest before commencement of the sprint protocol and immediately upon
131 completion of the sprint protocol, descriptions of collection and analysis is detailed in *Venous*
132 *blood sampling, eHsp72 and SOD measurement* below.

133 *Venous blood sampling, eHsp72 and SOD measurement:* In line with previous work in the
134 field (Gibson, et al., 2014) a 10 mL whole venous blood sample was drawn from the
135 antecubital fossa. Each sample was divided equally into 5 mL tubes (Starstedt, Germany)

136 containing EDTA (Vacuette®, Greiner BIO-one, UK). Whole blood samples were
137 centrifuged (Eppendorf 5804 R Centrifuge) at 4,500 rpm for 15 min to separate plasma.
138 Plasma was pipetted (Eppendorf Research/Research Pro) into 1.5 mL microtubes (Eppendorf)
139 and stored at -86°C (Sanyo Ultra Low, VIP Series) until analysis.

140 eHsp72 analysis utilised a commercially available pre-prepared Enzyme-Linked
141 Immunosorbent Assay (ELISA) kit in line with manufacturer's instructions (Stress Express
142 HSP70 High sensitivity ELISA kit, EKS-715, Stressgen Bioreagents, Victoria, Canada)
143 utilising a plate reader (ELx800, Bio-Tech Instruments, Inc. Winoski, USA) and read at an
144 absorption of 450 nm. The sensitivity of the ELISA kit was 0.09 ng/mL and both inter- and
145 intra-assay coefficient of variation was 3.2%, in line with previous work in the field (Gibson,
146 et al., 2014).

147 SOD was analysed with a commercially available ELISA kit (Cayman Chemical, Ann Arbor,
148 Michigan, USA) in line with manufacturer's instructions utilising a plate reader and read at
149 an absorption of 450 nm. The dynamic range of the SOD assay was 0.025-0.25 units/mL
150 SOD, and the inter- and intra-assay coefficients of variance were less than 3.7% (Cayman
151 Chemicals SOD ELISA kit, Cayman Chemical, USA).

152 *Plasma Osmolality:* Approximately 20µL of plasma was used to determine if changes in
153 plasma osmolality (Micro Osmometer Model 3300, Advanced Instruments, Inc., USA)
154 between conditions affected the final concentrations of eHsp72 and SOD. Changes in whole
155 venous plasma volume were quantified using established methods in triplicate (~50 µL) in
156 line with Gibson et al (2014). Plasma volume was not significantly different ($p=0.05$)
157 between any conditions.

158 *Supplementation:* Subjects consumed one pre-prepared 250mL bottle (70mL EM-X
159 (Effective Microorganisms UK) mixed with 180mL water) of EM-X drink, or, volume, taste
160 and odour matched 250mL bottle of placebo drink for seven consecutive days prior to

161 laboratory attendance. The EM-X drink was prepared in accordance with the manufacturer's
162 recommended dosage (personal communication).

163 Subjects reported to the laboratory on six occasions having fasted for 2 hours and replicating
164 food intake and activity levels (habitual exercise only) prior to each experimental visit.
165 Compliance for all the aforementioned experimental controls was at 100% in all subjects. The
166 protocol was approved by the institutional Ethics Committee and all subjects signed informed
167 consent following the principles outlined in the Declaration of Helsinki.

168 Experimental design – Visit 1

169 *Anthropometric measures:* During the initial visit subject's height and body mass were
170 obtained to the nearest 0.1cm/kg in the Frankfurt plane (Harpenden Instruments, West
171 Sussex, UK).

172 *VO_{2max} test:* Maximal aerobic capacity was determined using an incremental test to
173 exhaustion on a treadmill (Woodway, Waukesha, WI, USA). Participants began the test at a
174 speed of 9km.h⁻¹, 1% gradient. The test consisted of 2 minute stages, during the second
175 minute of each stage expired air was collected for approximately 60s via open circuit
176 spirometry. HR was monitored throughout the test using a HR monitor (Polar Sports Tester,
177 Polar, Electro Inc, Finland). Treadmill speed was increased by 1 km.h⁻¹ at the onset of each
178 new stage. Subjects were instructed to continue for as long as possible with verbal
179 encouragement throughout. VO_{2max} was taken as the highest VO₂ value obtained in any 10
180 second period and was taken as having been achieved when meeting end-point criteria in
181 accordance with the guidelines of the British Association of Sport and Exercise Sciences
182 (Bird & Davison, 1997). Expired air was analysed using an infrared and paramagnetic
183 analyser (model 1400, Servomex Controls, Crowborough, UK). Results of the VO_{2max} test
184 were used to calculate running speeds for the active recovery stage by using the linear
185 regression equation generated from the graph of running speed (km.h⁻¹) and VO₂ (mL.kg⁻¹
186 .min⁻¹).

187 Visit 2

188 *Maximal Anaerobic Running Test (MART)*: Two days after the VO_{2max} test, subjects began a
189 standardised warm up on a motorised treadmill (Woodway, Waukesha, WI, USA) and then
190 began the MART (Nummela, Alberts, Rijntjes, Luhtanen, & Rusko, 1996) at a speed of 12
191 $km.h^{-1}$, 10% gradient running for 20 seconds, runs were followed by 100 seconds of passive
192 recovery before beginning a new stage whereby speed was increased by $0.5 km.h^{-1}$. The test
193 was terminated when the subject could no longer keep pace with the treadmill, the final
194 completed running speed was used to prescribe the running velocity during the intermittent
195 running protocol.

196 Visits 3 to 6

197 Subjects completed one high intensity interval running protocol (HIT_{RP}) on each of the four
198 remaining visits in a randomised order utilising a double blind, cross over design, within a
199 purpose built environmental chamber with temperature and humidity controlled using
200 automated computer feedback (WatFlow control system; TISS, Hampshire, UK). Two visits
201 were completed in temperate conditions ($TEMP$; $20.4 \pm 1.7^{\circ}C$, $41, \pm 4.2 \% RH$), and two in
202 hot and humid conditions (HOT ; $34.7 \pm 2.0^{\circ}C$, $51.7 \pm 4.5 \% RH$). One hot and one temperate
203 trial were completed following a period of supplementation with EM-X ($TEMP_{EMX}$ or
204 HOT_{EMX}) or placebo ($TEMP_{PLA}$ or HOT_{PLA}). The HIT_{RP} were conducted in their entirety for
205 all participants in each of the four experimental trials.

206 **Statistical Methods**

207 All statistical analyses were completed using IBM SPSS Statistics 19 (SPSS Inc., Chicago,
208 IL). The normality of each dependent variable was checked using quantile-quantile (Q-Q)
209 plots and deemed plausible in each instance. The central tendency and dispersion of each
210 dependent variable are therefore reported as the mean (SD). The effects of Ingestion (placebo
211 vs. EM-X) and Temperature (temperate vs. hot) on the response of each dependent variable

212 over the twenty intervals (or pre- and post-intervention for eHsp72 and SOD) were
213 investigated using linear mixed models. The best fitting covariance structure for each model
214 was identified by minimising the Hurvich and Tsai's criterion. The assumptions of normally
215 distributed residuals around a mean of zero and constant variance were checked using Q-Q
216 plots and scatter plots and deemed plausible. Statistical significance was accepted as $p \leq 0.05$.

217 **Results**

218 Plasma eHsp72 increased from pre- to post-exercise ($F=62.1$, $p=0.001$), however, this time
219 effect was moderated by Temperature ($F=30.8$, $p=0.001$) see Fig.1. In TEMP eHsp72
220 increased by $0.8 \text{ ng} \cdot \text{mL}^{-1}$ (95% CI=0.3 to $1.2 \text{ ng} \cdot \text{mL}^{-1}$, $p=0.005$), compared to $1.5 \text{ ng} \cdot \text{mL}^{-1}$ in
221 HOT (95% CI=1.1 to $1.9 \text{ ng} \cdot \text{mL}^{-1}$, $p=0.001$). The main effects for Ingestion ($F=1.7$, $p=0.20$)
222 and the interaction effect ($F=0.3$, $p=0.59$) were not significant. The mean difference in
223 plasma eHsp72 between the placebo and EM-X conditions was not different in TEMP and
224 HOT ($F=0.4$, $p=0.53$).

225 SOD activity increased over time ($F=70.1$, $p=0.001$), the effect of which, was moderated by
226 Temperature ($F=9.4$, $p=0.004$) see Fig. 2. In TEMP there was a $0.10 \text{ U} \cdot \text{mL}^{-1}$ mean increase
227 in SOD activity (95% CI=0.047 to 0.16 units, $p=0.001$), compared to a $0.22 \text{ U} \cdot \text{mL}^{-1}$ mean
228 increase in HOT (95% CI=0.16 to $0.28 \text{ U} \cdot \text{mL}^{-1}$, $p=0.001$). There was no pre-intervention
229 difference between SOD activity in TEMP and HOT ($p=0.74$). SOD activity was higher in
230 HOT than TEMP post-intervention ($p=0.001$). Ingestion type did not influence SOD activity
231 over time, either alone ($F=1.7$, $p=0.20$), or as an interaction with Temperature ($F=1.4$,
232 $p=0.24$). Mean SOD was lower in the EM-X conditions compared to placebo ($F=9.7$,
233 $p=0.004$), but higher in HOT compared to TEMP ($F=6.7$, $p=0.014$). A significant interaction
234 was observed ($F=4.5$, $p=0.042$). In HOT there was a $0.10 \text{ U} \cdot \text{mL}^{-1}$ higher mean SOD activity
235 in the placebo condition compared to the EM-X condition (95% CI=0.045 to $0.16 \text{ U} \cdot \text{mL}^{-1}$,
236 $p=0.001$), with no difference between placebo and EM-X in TEMP (mean difference= 0.019
237 $\text{U} \cdot \text{mL}^{-1}$, 95% CI= -0.036 to $0.075 \text{ U} \cdot \text{mL}^{-1}$, $p=0.48$).

238 HR during the relief intervals increased by an average of $17 \text{ b}\cdot\text{min}^{-1}$ over the period of the 20
239 intervals ($F=491.7$, $p=0.001$), but, like T_c , this effect was not moderated by Ingestion
240 ($F=0.08$, $p=0.78$), but was moderated by Temperature ($F=10.9$, $p=0.001$) see Fig. 3. In
241 TEMP HR increased at a mean rate of $0.9 \text{ b}\cdot\text{min}^{-1}$ per interval, compared to $0.7 \text{ b}\cdot\text{min}^{-1}$ per
242 interval in HOT (mean slope difference= $0.2 \text{ b}\cdot\text{min}^{-1}$, 95% CI= 0.1 to $0.4 \text{ b}\cdot\text{min}^{-1}$, $p=0.001$).
243 This difference was due to an interaction effect, whereby the mean rate of increase in HR
244 decelerated at a mean rate of $0.07 \text{ b}\cdot\text{min}^{-1}$ per interval in HOT and $0.02 \text{ b}\cdot\text{min}^{-1}$ per interval in
245 TEMP (mean difference= $0.05 \text{ b}\cdot\text{min}^{-1}$, 95% CI= 0.02 to $0.07 \text{ b}\cdot\text{min}^{-1}$, $p=0.001$). Although the
246 rate of increase in HR during successive relief intervals was higher in TEMP, HR was, on
247 average, $14 \text{ b}\cdot\text{min}^{-1}$ higher in HOT than TEMP (95% CI= 13 to $15 \text{ b}\cdot\text{min}^{-1}$, $p=0.001$). The
248 main effect for Ingestion ($F=1.5$, $p=0.28$) and the interaction effect ($F=2.5$, $p=0.11$) were not
249 statistically significant.

250 T_c increased by $\sim 1.6^\circ\text{C}$ over the period of the 20 intervals ($F=163.8$, $p=0.001$). The rate of
251 increase was moderated by Temperature. In TEMP T_c increased at a mean rate of 0.062°C per
252 interval, compared to 0.081°C in HOT (mean slope difference= 0.019°C , 95% CI= 0.013 to
253 0.025°C , $p=0.001$). Mean T_c was, on average, 0.32°C higher in HOT compared to TEMP
254 (95% CI= 0.29 to 0.35°C , $p=0.001$). Although the main effect for Ingestion was not
255 significant ($F=2.3$, $p=0.19$), an interaction was observed ($F=18.4$, $p=0.001$). In TEMP no
256 difference in T_c was observed between placebo and EM-X (mean difference= 0.004°C , 95%
257 CI= -0.10 to 0.11°C , $p=0.93$), whereas in HOT mean T_c was 0.12°C higher for placebo
258 compared to EM-X (95% CI= 0.016 to 0.22°C , $p=0.03$).

259 TS increased by around 2 units during the 20 intervals ($F=143.9$, $p=0.001$), with the rate of
260 increase moderated by Temperature (Fig. 4). TS increased at a mean rate of 0.12 units per
261 interval in HOT and 0.09 units in TEMP (mean slope difference= 0.03 units, 95% CI= 0.02 to
262 0.04 units, $p=0.001$). There was no significant effect for Ingestion ($F=0.2$, $p=0.70$) and the
263 interaction also was not significant ($F=2.1$, $p=0.15$). However, an Ingestion-by-Temperature
264 interaction was observed ($F=13.3$, $p=0.001$), highlighting that in TEMP TS was 0.4 units

265 lower in the EM-X condition compared to placebo (95% CI=0.2 to 0.6 units, $p=0.006$),
266 whereas in HOT the 0.2 unit difference between placebo and EX--X did not reach statistical
267 significance (95% CI=-0.03 to 0.4 units, $p=0.08$).

268 RPE increased by around 7 units during the 20 intervals ($F=37.7$, $p=0.002$), with the rate of
269 increase moderated by Temperature (Fig. 4). The RPE increased at a rate of 0.35 units per
270 interval in HOT and by 0.28 units in TEMP (mean slope difference=0.06 units, 95% CI=0.04
271 to 0.09 units, $p=0.001$). The RPE was, on average, 0.7 units higher in HOT compared to the
272 TEMP (95% CI=0.6 to 0.9 units, $p=0.001$). The main effect for Ingestion ($F=3.6$, $p=0.12$)
273 and the interaction effect ($F=2.4$, $p=0.12$) were not statistically significant.

274 **Discussion**

275 eHsp72 and SOD concentrations increased pre to post exercise in HOT and TEMP (see Fig. 1
276 and Fig. 2). Elevated eHsp72 in HOT (+418%) has been reported elsewhere but not at the
277 magnitude to which our data observed (Whitham et al., 2007 (+200%); Magalhães et al.,
278 2010 (+34%); Periard et al., 2012 (~125%); Gibson et al., 2014 (172%)). Our observed
279 increases in TEMP (+212%) are less frequently observed but comparable with others
280 (Whitham et al., 2006 (+200%); Whitham et al., 2007 (+100%)). A minimum endogenous
281 criteria is required to increase eHsp72, this criteria – sufficient change in the absolute
282 ($\geq 38.5^{\circ}\text{C}$; (Amorim, Yamada, Robergs, Schneider, & Moseley, 2008)) and rate of T_c increase
283 ($1.6^{\circ}\text{C}\cdot\text{hr}^{-1}$ (Gibson et al. 2014); 2.0 & $2.5^{\circ}\text{C}\cdot\text{hr}^{-1}$ (Périard et al. 2012)), and significant (≥ 153
284 $\text{beats}\cdot\text{min}^{-1}$) sympathetic activity (Gibson, et al., 2014) can be achieved via exercise, a
285 thermal environment, or a combination of the two. Even in TEMP conditions our mean HR
286 responses of ≥ 150 $\text{beats}\cdot\text{min}^{-1}$ and final rectal temperatures of $\sim 38.5^{\circ}\text{C}$ meet the above
287 required endogenous criteria, and the proposed α -adrenergic stimulation (Johnson &
288 Fleshner, 2006). Greater eHsp72 concentration in HOT are likely a result of either greater
289 magnitude of HR (HOT >170 $\text{beats}\cdot\text{min}^{-1}$; TEMP >160 $\text{beats}\cdot\text{min}^{-1}$; Fig. 3) and/or T_c (HOT

290 >38.5°C; TEMP >38.0°C; Fig. 3) responses, or the earlier attainment of each stimuli, hence
291 greater duration above the previously stated minimum endogenous threshold.

292 SOD is an established biomarker of oxidative stress. Increased physiological strain (Fig. 3)
293 when exercising under hyperthermia vs normothermia (Lafrenz, Wingo, Ganio, & Cureton,
294 2008) led to greater concentrations in HOT (+14.7%) than TEMP (+6.8%; Fig. 2).
295 Amelioration in SOD increases within the HOT_{EMX} (+11.8%) condition compared to HOT_{PLA}
296 (+17.7%) is likely facilitated by the potent antioxidant capacity of EM-X. Amelioration was
297 not observable in TEMP where the difference between TEMP_{EMX} (+6.7%) and TEMP_{PLA}
298 (+6.9%) were negligible due to reduced physiological strain. EM-X composition is diverse
299 containing ~40 minerals and compounds, of which many have antioxidant effects both *in-*
300 *vitro* and *in vivo* (ubiquinone, α -tocopherol (vitamin E), lycopene, saponin and the flavonoid
301 quercetin (Aruoma, et al., 2002). We are unable to determine precisely which antioxidant
302 compound has potentially mediated this amelioration of oxidative stress, however, α -
303 tocopherol, lycopene, ubiquinone (Peternejl & Coombes, 2011) and flavonoids (Kressler,
304 Millard-Stafford, & Warren, 2011) are all known to exert an antioxidant influence *in vivo*
305 within humans. The use of a mixed antioxidant profile, rather than one specific antioxidant
306 compound, to ameliorate the reactive oxygen species response to exercise is supported
307 (Balakrishnan & Anuradha, 1998), giving efficacy to this ergogenic aid. Whilst reactive
308 oxygen species generation and/or disturbances to redox balance are essential components of
309 exercise adaption (Powers, et al., 2010), this is an undesirable response during repeated
310 competition due to the potential performance detriments (Cobley, et al., 2011). Ameliorating
311 redox balance disturbances can attenuate fatigue after repeated bouts of intermittent high-
312 intensity exercise within temperate environments (Cobley, et al., 2011).

313 Antioxidant supplementation during HIT related exercise performance, may be better suited
314 to tournament situations, whereby, fatigue “resistance/recovery from” is paramount and
315 adaptation is a negligible goal (Cobley, et al., 2011). Team sports require repeated sprint/HIT
316 based movement patterns, often in tournament situations in challenging environments, with

317 recovery time between competition suboptimal. EM-X supplementation elicited no such
318 negative side effects previously observed with N-Acetylcysteine supplementation (Cobley, et
319 al., 2011). Our data suggests the presence of reactive oxygen species, and the established
320 pathways which elicit decrements in repeated sprint exercise performance with limited
321 recovery time (Cobley, et al., 2011) appear to/could be attenuated by the supplementation of
322 EM-X during exercise in the heat.

323 No direct cause and effect relationship between EM-X supplementation and peripheral
324 mechanisms of exercise fatigue attenuation can be claimed from the present study. Future
325 studies should seek to quantify the potential ergogenic effect of EM-X, across a range of
326 relative dosages, on fatiguing high-intensity interval training exercise, within hot
327 environments akin to the study design of others (Cobley, et al., 2011). The efficacy of EM-X
328 as an ergogenic aid to attenuate oxidative stress resulting from prolonged continuous exercise
329 in the heat could additionally be considered as a means to ameliorate short-term negative
330 physiological effects which contribute to cumulative performance decrements observed
331 during consecutive multiday competition.

332 Classical physiological responses to exercise in hot vs. temperate conditions were observed,
333 with increased HR and T_c (Galloway & Maughan, 1997; Gibson, et al., 2014; Lafrenz, et al.,
334 2008), and TS and RPE (Gagge, et al., 1969; Galloway & Maughan, 1997; Gibson, et al.,
335 2014). Insufficient heat dissipation in HOT_{EMX} and HOT_{PLA} increased heat storage in
336 comparison with TEMP ($1.86^{\circ}\text{C}\cdot\text{hr}^{-1}$ in TEMP vs $2.43^{\circ}\text{C}\cdot\text{hr}^{-1}$ in HOT, see Fig. 3) irrespective
337 of the matched work, VO_2 and metabolic heat production (MHP). Increased physiological
338 (HR HOT >170 beats $\cdot\text{min}^{-1}$; TEMP >160 beats $\cdot\text{min}^{-1}$) and thermal responses (T_c HOT
339 $>38.5^{\circ}\text{C}$; TEMP $>38.0^{\circ}\text{C}$) during our HIT in comparison to data reported for continuous
340 exercise of approximately similar average intensity in the heat (Gibson, et al., 2014; Houmard
341 et al., 1990) are not unexpected as intermittent exercise is known to elicit greater thermal and
342 cardiovascular strain than continuous exercise of the same average intensity over a fixed
343 duration (Taylor and Cotter 2006). This difference is further increased when running in

344 comparison to cycling, where increased absolute VO₂ (Kang, Hoffman, Walker, Chaloupka,
345 & Utter, 2003), thus MHP, is also greater at a given % of VO_{2max} (Smoljanic, Morris, Dervis,
346 & Jay, 2014).

347 **Conclusion**

348 High intensity exercise in hot conditions elicited greater eHsp72 and SOD than matched
349 exercise performance in temperate conditions, likely due to the increases in relative exercise
350 intensity, and associated increases in physiological strain induced by hyperthermia.
351 Supplementation of EM-X attenuated the SOD increases in hot conditions, suggesting
352 oxidative stress had been reduced.

353 **Abbreviations**

354 eHsp72; Extracellular heat shock protein 72. EM-X; Effective Microorganism X. HIT; High-
355 intensity interval training. HIT_{RP}; High intensity interval running protocol. HOT; Hot and
356 humid conditions. HOT_{EMX}; Supplementation with EM-X in hot conditions. HOT_{PLA};
357 Supplementation with placebo in hot conditions. HR; Heart rate. iHsp72; Intracellular Hsp72.
358 MART; Maximal Anaerobic Running Test. RPE; Ratings of perceived exertion. SOD;
359 Superoxide dismutase. T_c; Core temperature. TEMP; Temperate conditions. TEMP_{EMX};
360 Supplementation with EM-X in temperate conditions. TEMP_{PLA}; Supplementation with
361 placebo in temperate conditions. TS; Thermal sensation. VO_{2max}; Maximum oxygen uptake

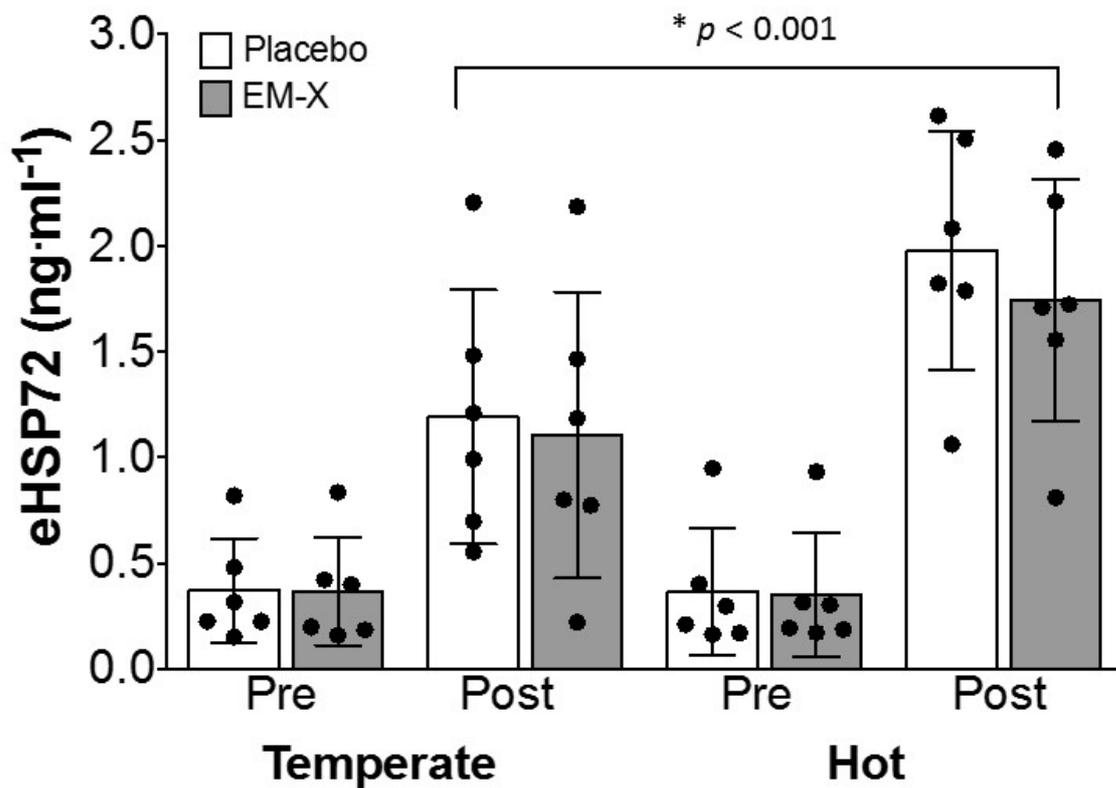
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363 **References**

- 364 Amorim, F. T., Yamada, P. M., Robergs, R. A., Schneider, S. M., & Moseley, P. L. (2008).
365 The effect of the rate of heat storage on serum heat shock protein 72 in humans.
366 *European Journal of Applied Physiology*, 104(6), 965-972.
- 367 Aruoma, O. I., Deiana, M., Rosa, A., Casu, V., Piga, R., Peccagnini, S., et al. (2002).
368 Assessment of the ability of the antioxidant cocktail-derived from fermentation of
369 plants with effective microorganisms (EM-X) to modulate oxidative damage in the
370 kidney and liver of rats in vivo: studies upon the profile of poly- and mono-
371 unsaturated fatty acids. *Toxicology Letters*, 135(3), 209-217.

- 372 Aruoma, O. I., Moncaster, J. A., Walsh, D. T., Gentleman, S. M., Ke, B., Liang, Y. F., et al.
373 (2003). The antioxidant cocktail, effective microorganism X (EM-X), protects retinal
374 neurons in rats against N-methyl-D-aspartate excitotoxicity in vivo. *Free Radical*
375 *Research*, 37(1), 91-97.
- 376 Ascensao, A., Rebelo, A., Oliveira, E., Marques, F., Pereira, L., & Magalhaes, J. (2008).
377 Biochemical impact of a soccer match - analysis of oxidative stress and muscle
378 damage markers throughout recovery. *Clinical Biochemistry*, 41(10-11), 841-851.
- 379 Balakrishnan, S. D., & Anuradha, C. V. (1998). Exercise, depletion of antioxidants and
380 antioxidant manipulation. *Cell Biochem Funct*, 16(4), 269-275.
- 381 Bird, S., & Davison, R. (1997). *Physiological Testing Guidelines*. Leeds: British Association
382 of Sport and Exercise Sciences.
- 383 Borg, G., Ljunggren, G., & Ceci, R. (1985). The increase of perceived exertion, aches and
384 pain in the legs, heart rate and blood lactate during exercise on a bicycle ergometer.
385 *Eur J Appl Physiol Occup Physiol*, 54(4), 343-349.
- 386 Cobley, J. N., McGlory, C., Morton, J. P., & Close, G. L. (2011). N-Acetylcysteine's
387 Attenuation of Fatigue After Repeated Bouts of Intermittent Exercise: Practical
388 Implications for Tournament Situations. *International Journal of Sport Nutrition and*
389 *Exercise Metabolism*, 21(6), 451-461.
- 390 Deiana, M., Dessi, M. A., Ke, B., Liang, Y. F., Higa, T., Gilmour, P. S., et al. (2002). The
391 antioxidant cocktail effective microorganism X (EM-X) inhibits oxidant-induced
392 interleukin-8 release and the peroxidation of phospholipids in vitro. *Biochemical and*
393 *Biophysical Research Communications*, 296(5), 1148-1151.
- 394 Do, J. S., Seo, H. J., Hwang, J. K., Kim, J. H., & Nam, S. Y. (2007). Effective microorganism
395 fermentation extract (EMA) attenuates airway hyperreactivity and inflammation
396 through selective inhibition of the TH2 response independently of antioxidant
397 activity. *International Journal of Molecular Medicine*, 20(4), 631-635.
- 398 Gagge, A. P., Stolwijk, J. A., & Saltin, B. (1969). Comfort and thermal sensations and
399 associated physiological responses during exercise at various ambient temperatures.
400 *Environ Res*, 2(3), 209-229.
- 401 Galloway, S. D., & Maughan, R. J. (1997). Effects of ambient temperature on the capacity to
402 perform prolonged cycle exercise in man. *Med Sci Sports Exerc*, 29(9), 1240-1249.
- 403 Gibson, O., Dennis, A., Parfitt, T., Taylor, L., Watt, P., & Maxwell, N. (2014). Extracellular
404 Hsp72 concentration relates to a minimum endogenous criteria during acute exercise-
405 heat exposure. *Cell Stress and Chaperones*, 19(3), 389-400.
- 406 Houmard, J. A., Costill, D. L., Davis, J. A., Mitchell, J. B., Pascoe, D. D., & Robergs, R. A.
407 (1990). The influence of exercise intensity on heat acclimation in trained subjects.
408 *Med Sci Sports Exerc*, 22(5), 615-620.
- 409 Johnson, J. D., & Fleshner, M. (2006). Releasing signals, secretory pathways, and immune
410 function of endogenous extracellular heat shock protein 72. *J Leukoc Biol*, 79(3), 425-
411 434.

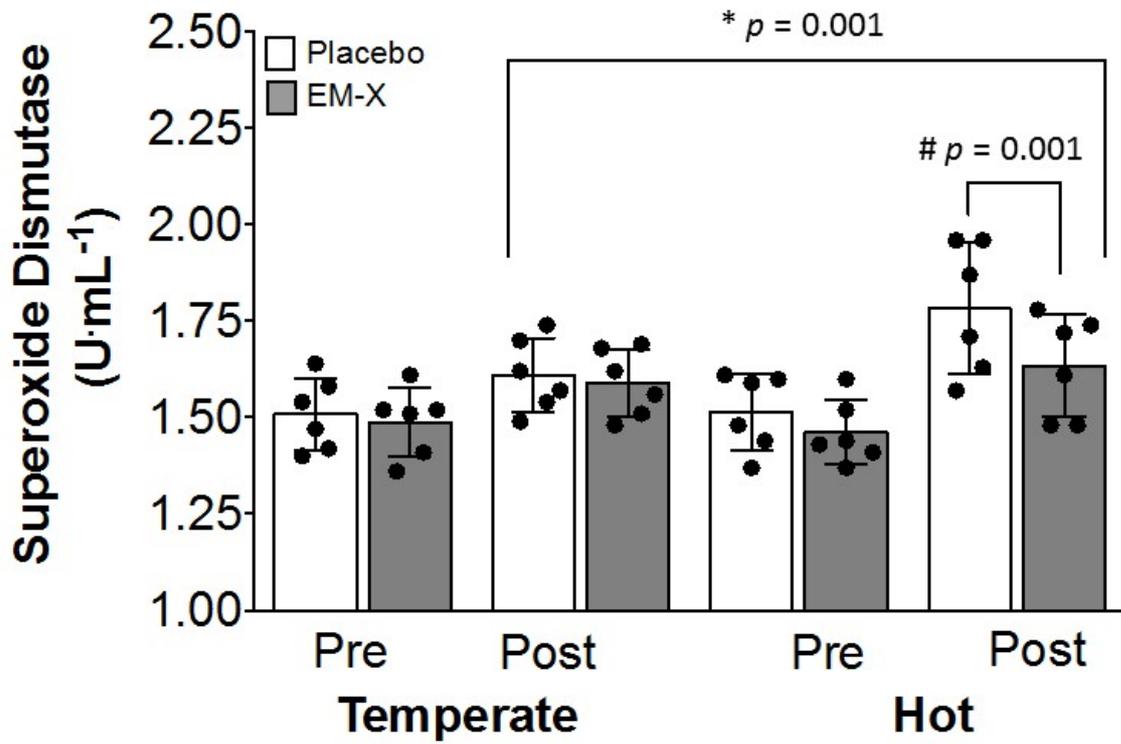
- 412 Kang, J., Hoffman, J. R., Walker, H., Chaloupka, E. C., & Utter, A. C. (2003). Regulating
413 intensity using perceived exertion during extended exercise periods. *Eur J Appl*
414 *Physiol*, 89(5), 475-482.
- 415 Ke, B., Liang, Y. F., Zhong, Z. X., Higa, T., & Aruoma, O. I. (2005). Evaluation of the
416 toxicity and safety of the antioxidant beverage effective microorganisms-X (EM-X) in
417 animal models. *Environmental Toxicology and Pharmacology*, 20(2), 313-320.
- 418 Kressler, J., Millard-Stafford, M., & Warren, G. L. (2011). Quercetin and Endurance Exercise
419 Capacity: A Systematic Review and Meta-analysis. *Medicine and Science in Sports*
420 *and Exercise*, 43(12), 2396-2404.
- 421 Lafrenz, A. J., Wingo, J. E., Ganio, M. S., & Cureton, K. J. (2008). Effect of ambient
422 temperature on cardiovascular drift and maximal oxygen uptake. *Med Sci Sports*
423 *Exerc*, 40(6), 1065-1071.
- 424 Nummela, A., Alberts, M., Rijntjes, R. P., Luhtanen, P., & Rusko, H. (1996). Reliability and
425 validity of the maximal anaerobic running test. [Article]. *International Journal of*
426 *Sports Medicine*, 17, S97-S102.
- 427 Periard, J. D., Ruell, P., Caillaud, C., & Thompson, M. W. (2012). Plasma Hsp72 (HSPA1A)
428 and Hsp27 (HSPB1) expression under heat stress: influence of exercise intensity. *Cell*
429 *Stress Chaperones*, 17(3), 375-383.
- 430 Peternelj, T.-T., & Coombes, J. S. (2011). Antioxidant Supplementation during Exercise
431 Training Beneficial or Detrimental? *Sports Medicine*, 41(12), 1043-1069.
- 432 Powers, S. K., Duarte, J., Kavazis, A. N., & Talbert, E. E. (2010). Reactive oxygen species
433 are signalling molecules for skeletal muscle adaptation. [Article]. *Experimental*
434 *Physiology*, 95(1), 1-9.
- 435 Reilly, T., Atkinson, G., Edwards, B., Waterhouse, J., Farrelly, K., & Fairhurst, E. (2007).
436 Diurnal variation in temperature, mental and physical performance, and tasks
437 specifically related to football (soccer). *Chronobiology International*, 24(3), 507-519.
- 438 Sawka, M. N., Burke, L., Eichner, R., Maughan, R., Montain, S. J., & Stachenfeld, N. (2007).
439 Exercise and fluid replacement. *Medicine & Science in Sports & Exercise*, 39(2), 377-
440 390.
- 441 Smoljanic, J., Morris, N. B., Dervis, S., & Jay, O. (2014). Running economy, not aerobic
442 fitness, independently alters thermoregulatory responses during treadmill running. *J*
443 *Appl Physiol*, 117(12):1451-9
- 444 Taylor, L., Hillman, A., Midgley, A., Peart, D., Christmas, B., & McNaughton, L. (2012).
445 Hypoxia mediated prior induction of monocyte expressed HSP72 and HSP32 provides
446 protection to the sub-maximal exercise induced disturbances to redox balance. *Amino*
447 *Acids*, 43(5), 1933-1944.
- 448 Winget, C. M., Deroshia, C. W., & Holley, D. C. (1985). Circadian-rhythms and athletic
449 performance. *Medicine and Science in Sports and Exercise*, 17(5), 498-516.
- 450 Yamada, P., Amorim, F., Moseley, P., & Schneider, S. (2008). Heat shock protein 72
451 response to exercise in humans. *Sports Medicine*, 38(9), 715-733.



453

454 **Figure 1.** Mean (SD) plasma eHsp72 concentrations at pre- and post-exercise for the four
 455 experimental conditions. Spots represent individual subject data * Post-exercise plasma
 456 eHsp72 in hot conditions significantly higher than post- exercise plasma eHsp72 in temperate
 457 conditions

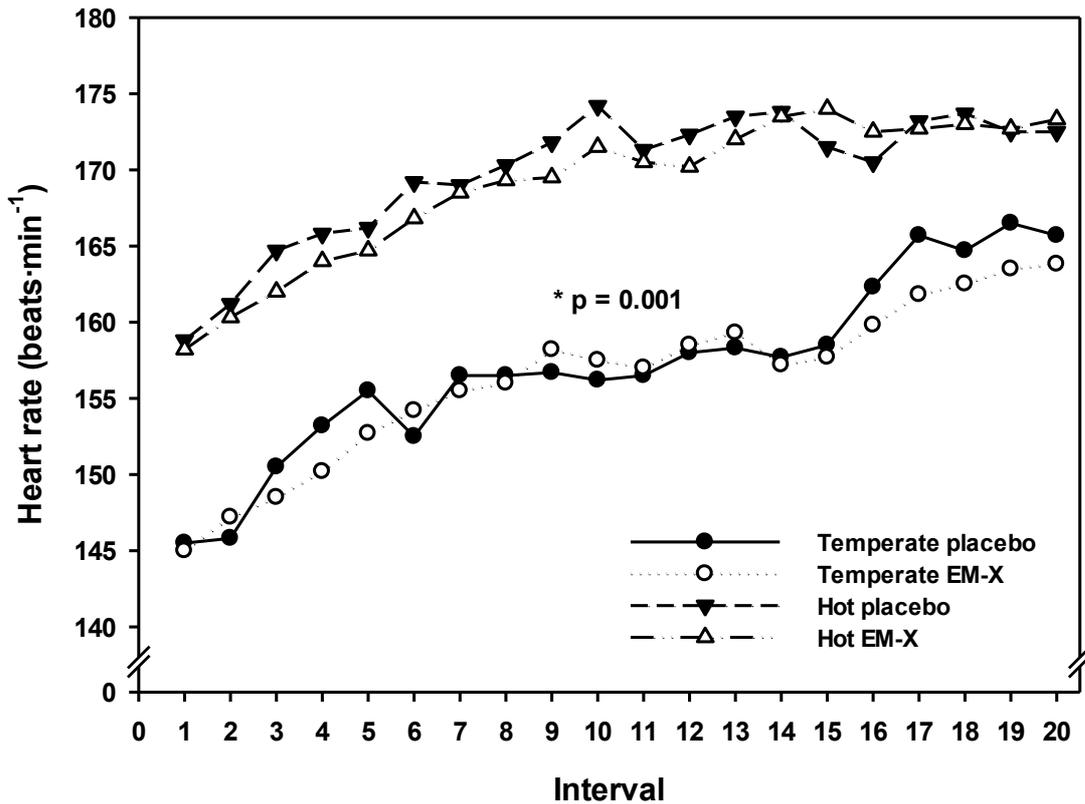
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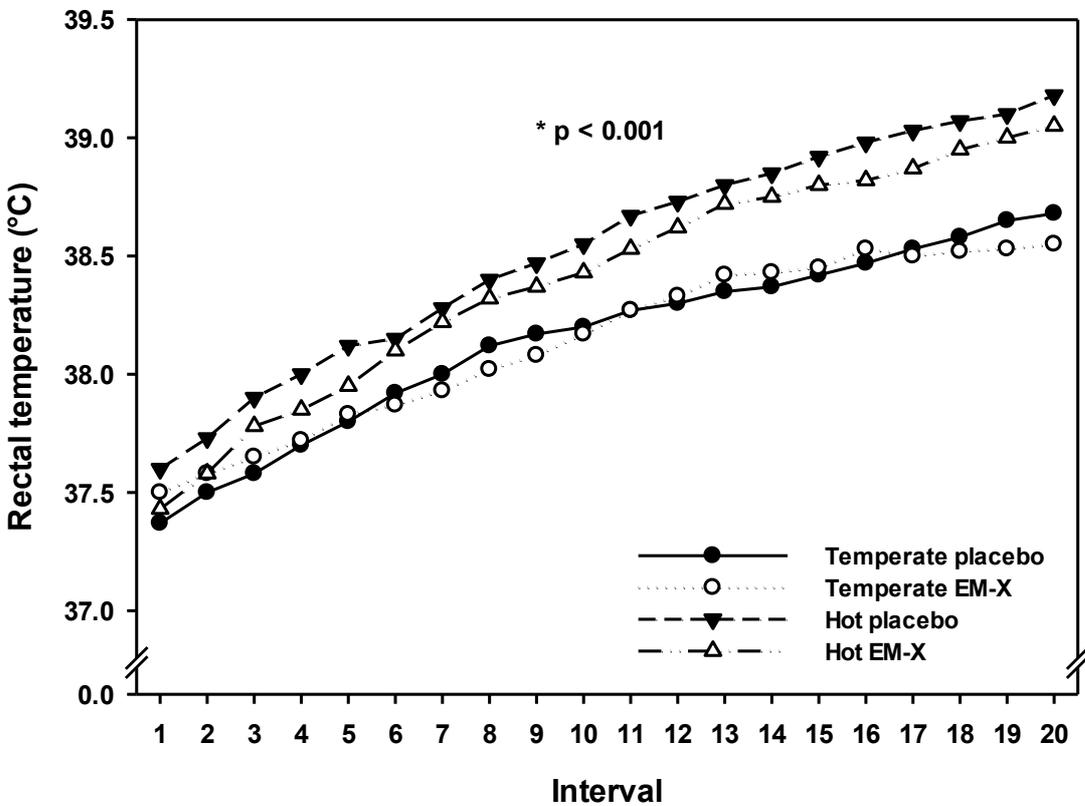
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460 **Figure 2.** Mean (SD) superoxide dismutase (SOD) concentrations at pre- and post-exercise
 461 for the four experimental conditions. Spots represent individual subject data. * Mean post-
 462 exercise SOD activity in hot conditions significantly higher than mean post-exercise SOD
 463 activity SOD activity in temperate conditions. # Mean post-exercise SOD activity lower
 464 following EM-X vs. Placebo in hot conditions.

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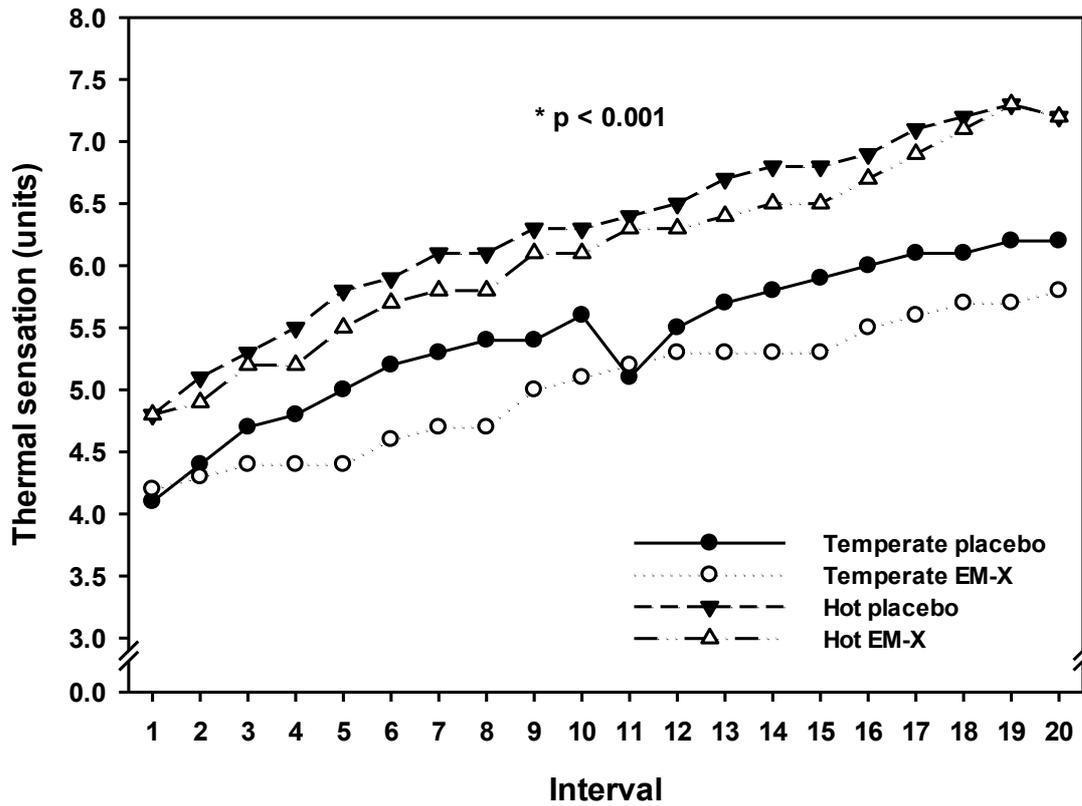


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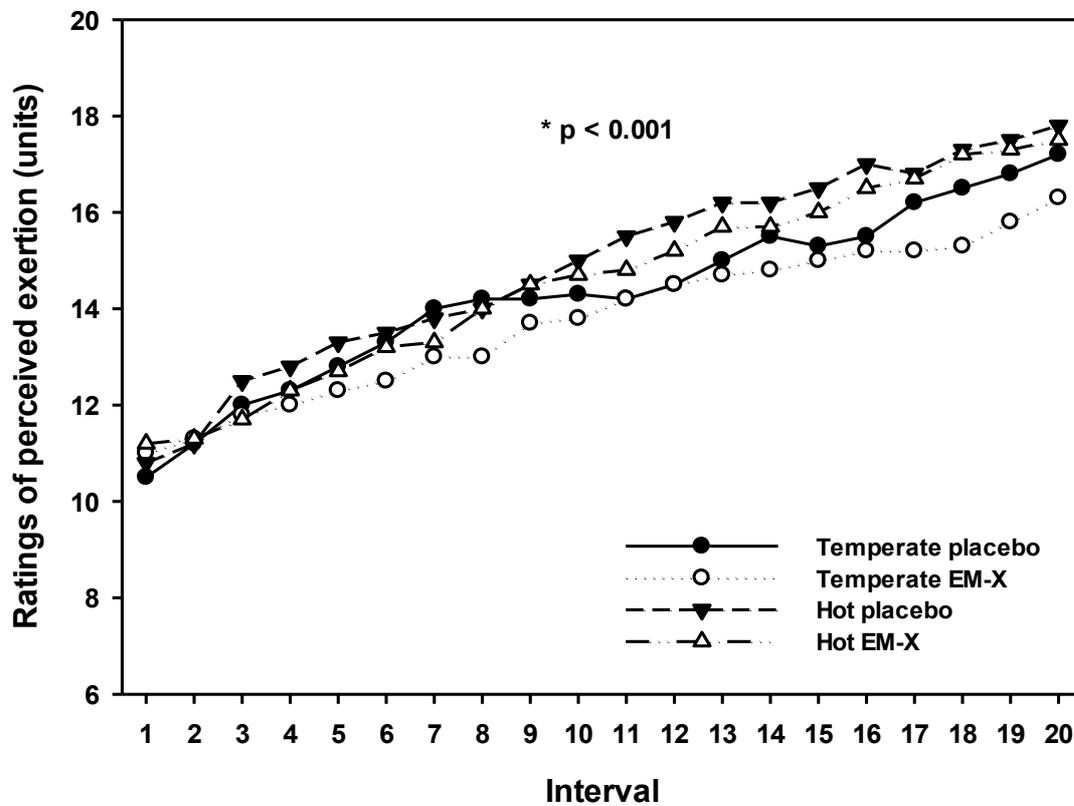


467

468 **Figure 3.** Physiological responses (Mean HR; top. T_c ; bottom) over the 20 intervals in the
 469 four experimental conditions. Error bars have been omitted for clarity. * Slopes for hot
 470 conditions significantly higher than for the temperate conditions.



471



472

473 **Figure 4.** Perceptual responses (Mean TS; top. RPE; bottom) over the 20 intervals in the four
 474 experimental conditions. Error bars have been omitted for clarity. * Slopes for hot conditions
 475 significantly higher than for the temperate conditions.